



Enhanced and sustained transdermal delivery of primaquine from polymeric thermoresponsive hydrogels in combination with Dermarollers®

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ABSTRACT

Primaquine (PMQ) is an effective antimalaria drug with several limitations. We report the combinatorial approach of thermoresponsive hydrogels and Dermarollers® for transdermal delivery of PMQ to overcome these limitations. The hydrogels were prepared using Pluronic F127 (PF127) and F68 (PF68). Specifically, HPMC was added into the formulation to improve the bioadhesive properties. Numerous formulations were prepared, showing that formulation comprising 15 % PF127, 3 % PF68 and 0.4 % HPMC with 1 % PMQ was selected as the optimum formulation. The formulation showed the gelation temperature around 35 °C with bioadhesive strength of 26.43 ± 2.31 dyne.cm². Importantly, the pH of the formulation was suitable for skin application with the percentage of PMQ recovery of 99.57 ± 3.23 %. Moreover, the hydrogels exhibited free-flow liquid at storage and room temperature and high viscosities in the skin temperature. *In vitro* release experiments showed that the release of PMQ was sustained for 24 h. Evaluated in extensive *ex vivo* studies, the treatment with Dermarollers® improved the skin permeation and retention of PMQ for 3 days. In combination with Dermarollers®, the *ex vivo* permeation of PMQ was sustained and the localization of PMQ in the skin was improved over 72 h.

1. Introduction

Around 228 million people in the world were affected by malaria in 2018 [1]. To overcome this, significant improvements have been made to decrease malaria-associated morbidity and death [2,3]. Primaquine (PMQ) has been established as an effective antimalaria drug [4]. Nevertheless, the effectiveness of PMQ is hindered by various problems. Following the administration orally, PMQ experiences first-pass metabolism in the liver, forming carboxyprimaquine (cPQ) through oxidative deamination. This metabolite has been found to show less antimalarial effect and high haemotoxic effect. Additionally, other side effects have been also reported after this administration, including abdominal pain, bitter taste, nausea and long therapy duration [4,5].

Being an attractive route, transdermal delivery has been a promising alternative to oral administration. The delivery of substances transdermally exhibits numerous benefits, namely avoiding first-pass metabolism, painless, excellent compliance and easy accessible [6–8]. Accordingly, this route could be considered as an alternative route to deliver PMQ. However, because of the specific compositions comprised in the skin, the delivery of drugs through this route is challenging [8].

The presence of lipophilic layer, *stratum corneum* (SC), has become the foremost physical blockade for drug permeated through the skin [9]. Moreover, PMQ shows low permeability. Microneedles are alternative devices for transdermal delivery which can disturb SC and create pores, enabling the delivery of drugs to the skin [10–12]. Dermarollers® are one of sorts of microneedles, belonging to solid microneedle type, which have been widely utilized in the cosmetic applications [9,13]. A plethora of research explored the effectiveness of Dermarollers® to enhance the pharmacology effect of numerous active pharmaceutical ingredients [13–15].

It is also important to consider the dosage form of PMQ for this purpose. The poor adherence of the patient is challenge in the malaria therapy. Accordingly, it is important to develop a delivery system that can sustain the release of PMQ through the skin. Thermoresponsive hydrogels are able to turn from a liquid to a gel when moved from the room temperature to the body temperature [16]. This system has been extensively applied as *in situ* forming vehicle because the sustain release behavior. Several polymers have been explored to prepare this approach. Amongst numerous polymers, in this study, Pluronic® was used as the thermoresponsive agent [17]. The properties of this polymer

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are sensitive to temperatures, making it suitable for the development of thermosensitive hydrogels with various applications. Importantly, Pluronic® is compatible with several type of cells and biological fluids, as well as non-irritant [16,18,19]. To improve its effectiveness, bioadhesive agents can be included into thermoresponsive formulations. Hydroxy propyl methyl cellulose (HPMC) has been recognized to show this property [20]. It has been reported that, combined with micro-needles, this system has successfully improved the penetrability of fluorescein sodium [21]. However, there has been no study reported using the therapeutic agents. Therefore, in this study, for the first time, we developed thermoresponsive hydrogel formulations with bioadhesive properties containing PMQ. Here, we reported the improved transdermal delivery of PMQ using the combination of thermoresponsive and Dermarollers®. Not only that, the deposition of PMQ in the skin was also improved by this combinatorial approach.

2. Material and methods

2.1. Materials

Primaquine biphosphate (PMQ) (purity, $\geq 98\%$) was purchased from Sigma Aldrich (Singapore). Pluronic® F68 (PF68) and Pluronic® F127 (PF127) were kindly gifted by BASF SE (Jakarta, Indonesia). Solid micro-needles (Dermarollers®) were obtained from SQY® (Guangdong, China).

2.2. Formulation of thermoresponsive *in situ* gel of primaquine

The cold method was used to fabricate the thermoresponsive hydrogels [22]. The composition of the hydrogels is depicted in Table 1. Initially, PF68 and PF127 were dissolved in cold distilled water (4 °C) using constant mixing until forming clear solution. Afterwards, HPMC and PMQ were added into the polymeric solution and stored in the refrigerator, forming clear solutions.

2.3. Determination of gelation temperature and bioadhesive properties

A test tube inverting technique in a water bath was applied to determine the gelation temperature of the formulation [23]. The tube containing the formulation was placed at 20 °C. The temperature was steadily increased until 55 °C. The gelation temperature was denoted when the solution turned into a gel after turning the tube over to 90° for 30 s. To determine the bioadhesion strength in skin tissue, a modified physical balance was applied [16]. In this study, an abdomen part of rats' skin was used. The skin was shaved and washed using distilled water. Prior to the experiment, the formulations were placed at 37 °C for 10 min and then applied to the skin attached to the balance in one side. Afterward, weights were placed in the other side of the balance until the formulations were detached from the skin. Finally, the bioadhesive strength was determined using the following calculation:

Table 1

The composition of thermoresponsive hydrogels containing PMQ (% w/v).

Formula	PMQ	PF127	PF68	HPMC
F1	1	18	–	–
F2	1	16	2	–
F3	1	15	3	–
F4	1	14	4	–
F5	1	10	8	–
F6	1	15	3	0.20
F7	1	15	3	0.40
F8	1	15	3	0.60
F9	1	15	3	0.80
F10	1	15	3	1

$$\text{Bioadhesive strength} = \frac{\text{weight (g)} \times 100}{\text{surface area (cm}^2\text{)}} \times 9.81$$

2.4. Determination of pH, viscosity and rheological behavior

The pH of the hydrogel formulation was determined using pH meter. The viscosity and rheology thermoresponsive gel were measured using a DV-III viscometer (RV model, Brookfield, USA). For viscosity measurement, gels were tested at cold temperature (4 °C), room temperature (25 °C) and skin temperature (32 °C).

2.5. Drug content analysis and *in vitro* release study of PMQ

Drug content of PMQ was assessed using spectrophotometer UV-Vis at 265 nm. The *in vitro* release of PMQ from thermoresponsive hydrogel was carried out using membraneless dissolution method using phosphate-buffered saline (PBS) as release media. Briefly, 10 g of hydrogel was placed into orbital shaker at 37 ± 0.5 °C. After the formulation became gel, PBS (5 mL) was added into the gels. At predetermined time (0.25 h, 0.5 h, 0.75 h, 1 h, 2 h, 3 h, 4 h, 5 h, 6 h, 7 h, 8 h, 12 h and 24 h), 1 mL of medium was taken and replaced with fresh medium. The concentration of PMQ was assessed using spectrophotometer UV-Vis at 265 nm.

2.6. *Ex vivo* skin permeation and retention study of PMQ

The *ex vivo* permeation of PMQ from thermoresponsive hydrogels was performed through rats' skin using vertical Franz diffusion cell with PBS as media. The skin was placed between donor and receiver compartment of Franz diffusion cell. The diffusion cell was stirred at 100 rpm and the temperature was maintained at 37 ± 0.5 °C. The formulation (equal 10 mg of PMQ) was placed into donor compartment. With total time of 72 h, at predetermined time, 0.5 mL of permeation media was withdrawn and replaced with new media. In this study, the effect of skin pretreatment with various length of Dermarollers®, namely 0.5 mm, 1 mm and 1.5 mm, was evaluated. Moreover, the concentration of PMQ retained in the skin was determined by extracting PMQ from the skin using methanol and analyzed using spectrophotometer UV-Vis at 265 nm.

3. Results and discussion

3.1. Preparation of primaquine-loaded thermoresponsive *in situ* gel

The main purpose of this study was to overcome the delivery issue of PMQ in the malaria treatment. As discussed earlier, to sustain the release of PMQ transdermally, thermoresponsive hydrogels were developed. Pluronics were selected as the main polymer to form this approach. The representative images of the formulation are depicted in Fig. 1A and B.

These compounds are triblock copolymers containing poly (ethylene oxide)-b-poly (propylene oxide)-b-poly (ethylene oxide) (PEO-PPO-PEO) compositions. Due to the presence of PEO as hydrophilic part and PPO as hydrophobic part, these polymers possess amphiphilic characteristics. At low temperature, Pluronics are the form of solution because they are in the unimers form. When the temperature increases, the unimers connects each other, forming micelles and turning into semi-solid form. Here, we evaluated different concentration of PF127 and PF68 to obtain the suitable gelation temperature. The addition PF68 into PF127 has been found to show numerous advantages, particularly in improving the low gelation temperature of PF127 [19]. Several studies have combined other types of Pluronics into PF127 to improve the thermosensitive properties of the polymers [24,25]. Despite several benefits, it was found that Pluronics do not possess adequate bioadhesive properties [19]. Accordingly, to improve the adhesion to the skin, HPMC was used. HPMC has been reported to form an adequate

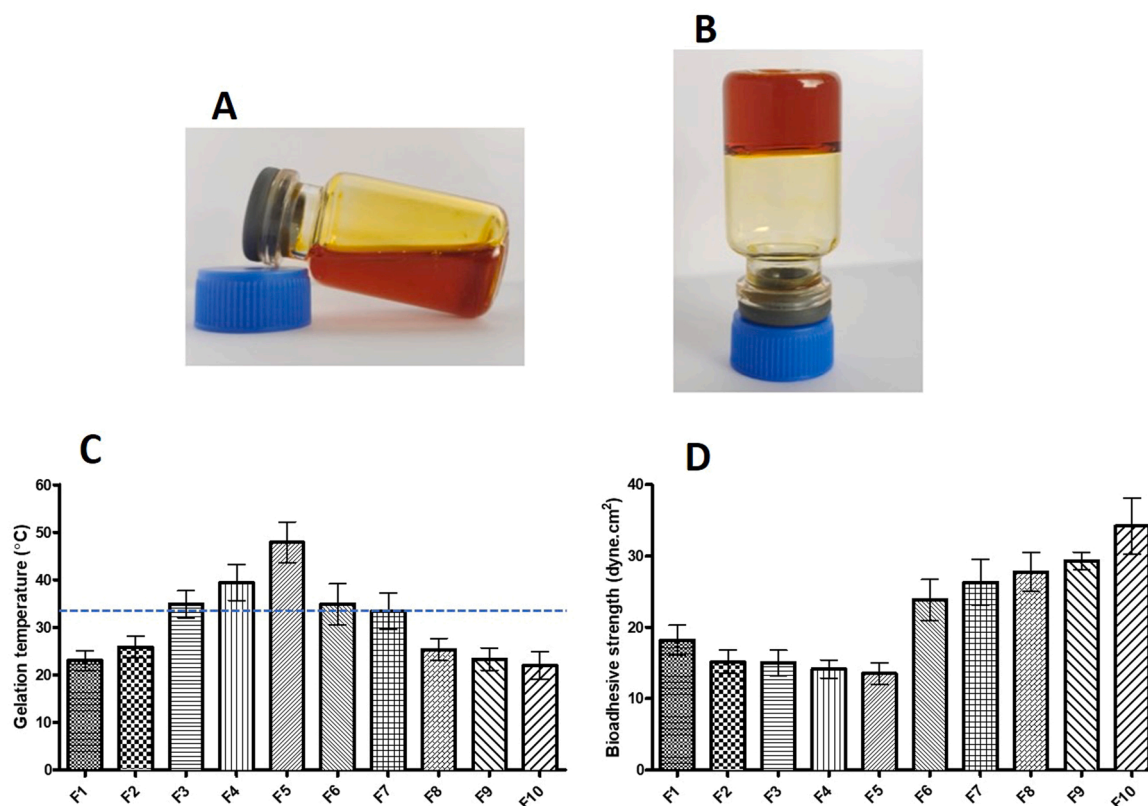


Fig. 1. Illustrative photograph of the formulation at room temperature (A) and skin temperature (B). The results of the determination of the gelation temperature of thermoresponsive hydrogels containing PMQ (mean \pm S.D., $n = 3$). The temperature of the skin is indicated by the dash blue line (C). The bioadhesion properties thermoresponsive hydrogels containing PMQ (mean \pm S.D., $n = 3$) (D). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

hydroxyl interaction with biological membrane, resulting in the strong bioadhesive property [18].

3.2. Determination of gelation temperature measurement

The ration of PPO/PEO of pluronics have been reported to affect the gelation temperature of the hydrogels. The hydrophobicity of PPO could decrease the gelation temperature, where the hydrophilicity of PEO could increase the gelation temperature. As PF127 contains more PPO than PF68, as presented in Fig. 1A, the higher concentration of PF68 resulted in lower gelation temperature. The use of PF127 alone resulted in the gelation temperature below the body temperature. This phenomenon was also shown in other studies [18,24,25], indicating the need of combining PF127 with other type of Pluronics. Our results showed that F3 containing 15 % PF127 and 3 % PF68 showed the gelation temperature around body temperature and, therefore, this ratio was investigated to evaluate the effect of HPMC as bioadhesive agent. As shown in Fig. 1C, above 0.4 %, the formulations possessed low gelation temperature (<32 °C), making the inappropriate for thermoresponsive preparations. Therefore, it was crucial to consider the consider the bioadhesive agent in the thermoresponsive preparations. It was important to ensure that the addition of bioadhesive compounds did not alter the thermosensitive properties of the preparations.

3.3. Determination of bioadhesion strength

To improve the adhesion with the desired site of action, hydrophilic polymers, HPMC, was added into the formulation. Fig. 1D shows that the bioadhesion strength of the hydrogels to the skin was improved following the increase on HPMC concentration. Interestingly, there were no significant differences ($p > 0.05$) in bioadhesive strength when

HPMC used in the concentration of 0.4 % compared to 0.6 % and 08 %. Although 1 % of HPMC showed significant improvement ($p < 0.05$) in bioadhesive properties, the formulation did not meet the requirement for thermosensitive preparations. In addition to skin delivery, HPMC has been widely used as bioadhesive agent for several application, including vaginal, ocular and oral [18,19,26], showing that the incorporation of HPMC in pharmaceutical dosage forms could potentially improve the bioadhesive properties the preparation in various biological membrane.

3.4. Determination of pH, viscosity and rheological behavior

It was crucial to ensure that the application of the hydrogels did not produce possible irritation to the skin. This could be achieved by preparing the formulation having pH around to the skin pH (± 5.8) [20]. The results exhibited that all hydrogels possessed pH values tolerated by the skin (Fig. 2A). Therefore, the application of the thermoresponsive hydrogels could not potentially cause any irritation of the skin.

The desired hydrogels were those which showed free-flow liquid with low viscosities at storage and room temperature and high viscosities in semisolid form in the skin temperature. Fig. 2B depicts that the ratio of PF127 and PF168 affected the viscosities of the hydrogels. Higher concentration of PF127 showed higher viscosity due to higher amount of triblock chains and, therefore, resulted in higher micelle size [25]. Showing similar trend with gelation temperature, the use of HPMC increased the viscosities of the hydrogels at all temperatures. As presented in Fig. 2B, only formulations containing HPMC with the concentrations below 0.6 % showed lower viscosities at room temperatures. Accordingly, F7 (0.4 % HPMC) was considered as the optimum formulation with appropriate properties for the thermoresponsive preparations. With respect to rheological properties, all formulations showed shear thinning behavior showing low viscosity in the high rate of share

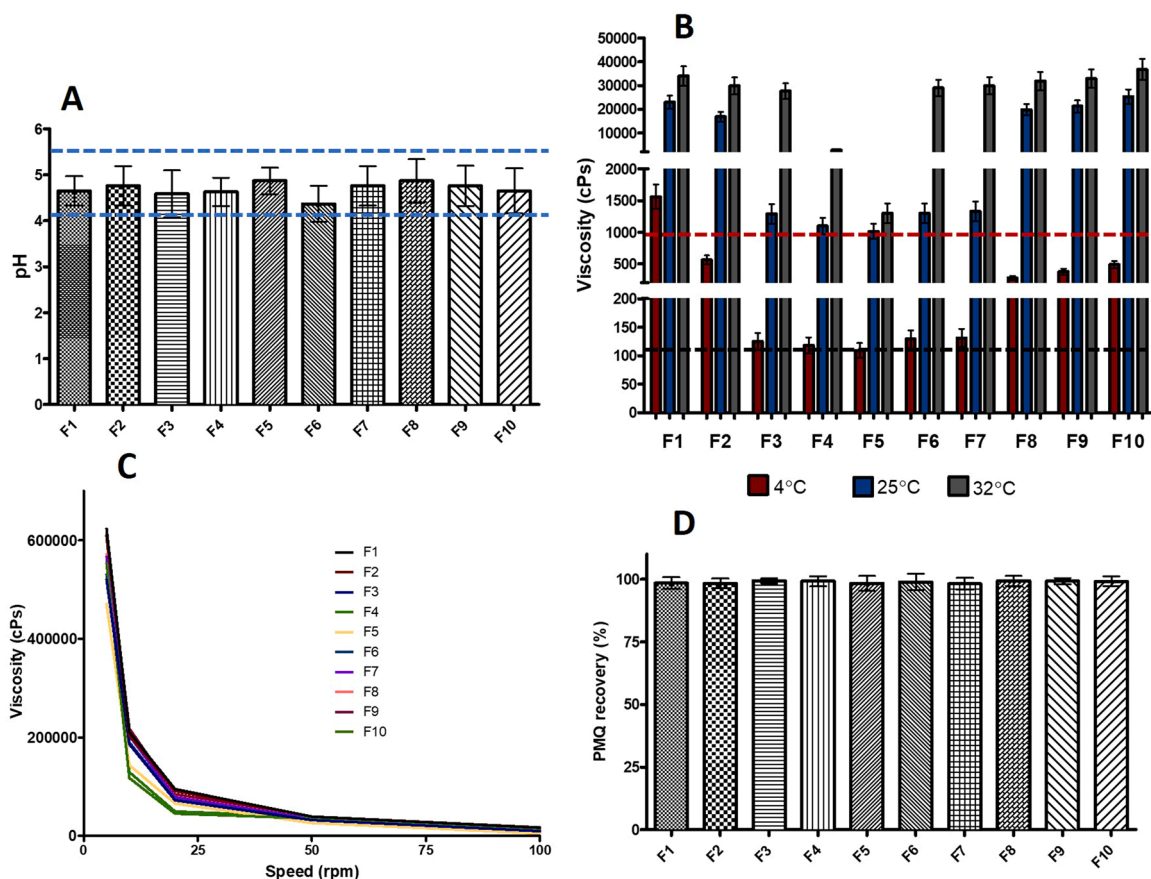


Fig. 2. The results of the pH determination of thermoresponsive hydrogels containing PMQ (mean \pm S.D., $n = 3$). The pH tolerated by the skin is indicated by the blue das lines (A). Viscosities of thermoresponsive hydrogels containing PMQ at various temperatures, including 4 °C, 25 °C, and 32 °C (mean \pm S.D., $n = 3$). The liquid form of the formulations is indicated by the values below black dash line and between black and red lines. The gel formation is indicated by the values above red dash line (B). The rheology pattern of thermoresponsive hydrogels containing PMQ (C). Drug recoveries percentages of thermoresponsive hydrogels containing PMQ (D) (mean \pm S.D., $n = 3$).

(Fig. 2C). These results were in good agreement with the gelation temperature determination, showing that the formulations with low gelation temperature exhibited higher viscosities at all temperature tested.

3.5. Drug content analysis and *in vitro* release study of PMQ

The concentration of PMQ in all formulations were between 98 % and 100 % (Fig. 2D). This indicated that the formulation process did not influence the concentration of PMQ in the formulation. It was previously reported that the percentage recovery of dosage forms should be between 95 % and 105 % [27]. In *in vitro* release, PMQ released exhibited biphasic manner. The *in vitro* release results of all formulations are shown in Fig. 3A and B. During the first hour, the burst release behavior was observed in all cases which might be due to the presence of PMQ on the surface of the gels. The burst release was previously shown in similar approach using fluorescein sodium as a model drug [21]. Furthermore, the sustained release was observed over 24 h, showing the ability of pluronics to sustain the release. It is important to note that F4 and F5 showing low gelation temperature, due to the liquid form at skin temperature, all PMQ released less than 5 h. Importantly, F7 which contained 0.4 % HPMC and showed desirable mucoadhesive and thermosensitive properties, exhibited similar behavior with optimum formulation without HPMC (F3). Accordingly, this formulation was selected for *ex vivo* studies.

3.6. *Ex vivo* skin permeation and retention study of PMQ

It was shown that due to the hydrophilicity of PMQ, it was difficult to

permeate the lipophilic of SC and the concentration of PMQ permeating the skin was less 25 % over 72 h. After the treatment using Dermarollers®, the concentration of PMQ permeating the skin was significantly enhanced with more than 50 % for 0.5 mm Dermarollers® and more than 75 % for Dermarollers® 1 mm and 1.5 mm. This was because the micropore created in the skin by Dermarollers®, disrupting the physical barrier of SC. It was important to note that sustained release behavior was observed over 72 h in all types of Dermarollers®. Interestingly, in *ex vivo* retention studies, it was found that the depositions of PMQ were significantly higher ($p < 0.05$) after the treatment using Dermarollers®. Considering non-significant different between 1 mm and 1.5 mm Dermarollers®, 1 mm was considered as the optimum length as the use of long needle could potentially cause discomfort of patient following its application. In our study, for the first time, it was concluded that the delivery of PMQ could be controlled using thermoresponsive hydrogels and Dermarollers®, providing sustained release permeation manner over 72 h, where PMQ was well-deposited in the skin and were released from thermoresponsive hydrogels. Several studies have investigated the administration of Dermarollers® to enhance the skin permeation of different drugs [13,14,28], showing the improvement of transdermal delivery in comparison with the conventional delivery systems. Previously, PMQ was developed in nanoemulsion delivery system [4]. It was reported that the nanoemulsion could improve and control the transdermal delivery for 24 h. In our study, the sustained release profile was maintained over 72 h. This was due to the formation of *in situ* depot from thermoresponsive system which allowed the improvement of control release pattern, indicating the inventiveness of our approach. However, before this approach can

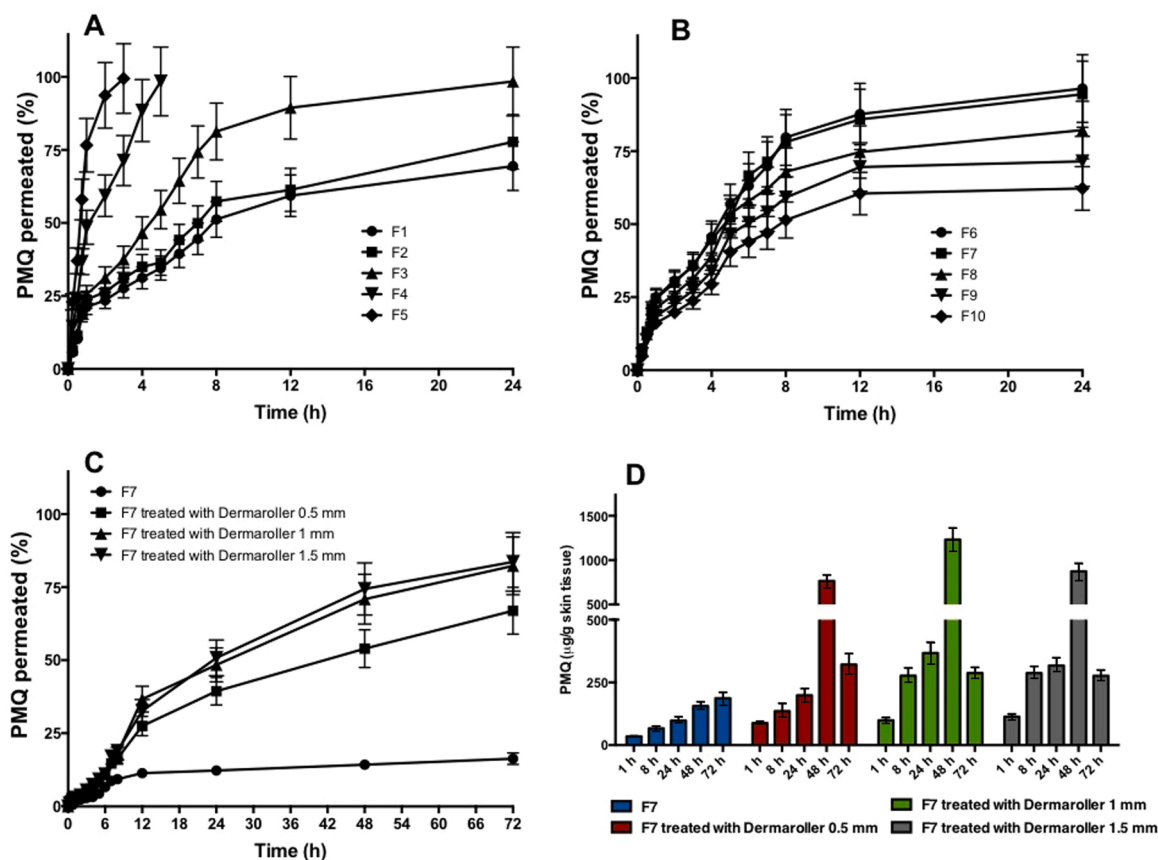


Fig. 3. *In vitro* release of PMQ from various thermoresponsive hydrogels (A and B) (mean \pm S.D., $n = 3$). *Ex vivo* skin permeation (C) and skin retention (D) of F7 as optimized formulation following the administration thermoresponsive hydrogels with and without the treatment of Dermarollers® with various length (mean \pm S.D., $n = 3$).

applied in the clinical treatment, several considerations are required. The sterility of this system should be considered. With regard to this, previous study has shown that the penetration of microorganism across the epidermis following the application of microneedles was found to be negligible [29]. Therefore, we could assume that the pores created by the use of microneedle should not cause the risk of infection. However, further studies are required to ensure this in the clinical applications. Moreover, the storage stability of the thermoresponsive gel should be investigated. It is also critical to ensure the reproducibility of this combination approach performed by patients. To achieve this, the pressure applied should be similar. Automated dermaroller has been developed to ensure the reproducibility of pressure applied [30], which could be potentially combined with the thermoresponsive developed in this study. Following this promising results, further *in vivo* studies should be carried out to investigate the efficacy, the pharmacokinetic profile and the dose determination of this approach. Moving forward, the impact of the deposition of the polymers should be evaluated. The use of Pluronics-based in the skin administration following microneedle applications is still limited. Therefore, the attention should not only be given to the pharmacokinetic profiles of the drugs, but also to the polymers used, particularly in the clearance profile. Finally, the scale-up manufacturing process should be considered in order to ensure that the method preparation can be applied in the industrial scale.

4. Conclusion

In this study, for the first time, we developed thermoresponsive hydrogels with bioadhesive properties to transdermally delivery PMQ combined with Dermarollers®. The combination of PF127 and PF68 with the ratio of 15 % and 3 % with 0.4 % HPMC was found to be the

optimal hydrogel with suitable thermoresponsive and mucoadhesive characteristics. The approach could potentially sustain the release of PMQ over 24 h. Essentially, the combination with Dermarollers® was able to enhance and control the *ex vivo* permeation and retention of PMQ through rats' skin over 72 h.

CRediT authorship contribution statement

Andi Dian Permana: Conceptualization, Methodology, Funding acquisition, Resources, Validation, Supervision, Writing – original draft. **Diany Elim:** Conceptualization, Data curation, Methodology, Funding acquisition, Resources. **Putri Wulandari Resky Ananda:** Methodology, Formal analysis, Investigation, Visualization. **Hilman Syamami Zaman:** Methodology, Investigation, Data curation. **Wahdaniyah Muslimin:** Data curation, Software, Validation. **Muhamad Gilang Ramadhan Tunggeng:** Methodology, Investigation.

Authors contribution

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

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Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data Availability

The authors do not have permission to share data.

References

- C.E. Umeyor, I. Okoye, E. Uronnachi, T. Okeke, F. Kenechukwu, A. Attama, Repositioning miconazole nitrate for malaria: formulation of sustained release nanostructured lipid carriers, structure characterization and in vivo antimalarial evaluation, *J. Drug Deliv. Sci. Technol.* 61 (2021), 102125, <https://doi.org/10.1016/j.jddst.2020.102125>.
- X. Dong, X. Zhang, M. Wang, L. Gu, J. Li, M. Gong, Heparin-decorated nanostructured lipid carriers of artemether-protoporphyrin IX-transferrin combination for therapy of malaria, *Int. J. Pharm.* 605 (2021), 120813, <https://doi.org/10.1016/j.ijpharm.2021.120813>.
- A. Kumari, M. Karnatak, D. Singh, R. Shankar, J.L. Jat, S. Sharma, D. Yadav, R. Shrivastava, V.P. Verma, Current scenario of artemisinin and its analogues for antimalarial activity, *Eur. J. Med. Chem.* 163 (2019) 804–829, <https://doi.org/10.1016/j.ejmech.2018.12.007>.
- P. Sharma, M. Tailang, Design, optimization, and evaluation of hydrogel of primaquine loaded nanoemulsion for malaria therapy, *Futur. J. Pharm. Sci.* 6 (2020), <https://doi.org/10.1186/s43094-020-00035-z>.
- G. Mihaly, S. Ward, G. Edwards, D. Nicholl, M. Orme, A. Breckenridge, Pharmacokinetics of primaquine in man. I. Studies of the absolute bioavailability and effects of dose size, *Br. J. Clin. Pharmacol.* 19 (1985) 745–750, <https://doi.org/10.1111/j.1365-2125.1985.tb02709.x>.
- S. Wiedersberg, R.H. Guy, Transdermal drug delivery: 30 + years of war and still fighting!, *J. Control. Release* 190 (2014) 150–156.
- T.M. Tuan-Mahmood, M.T.C. McCrudden, B.M. Torrisi, E. McAlister, M.J. Garland, T.R.R. Singh, R.F. Donnelly, Microneedles for intradermal and transdermal drug delivery, *Eur. J. Pharm. Sci.* 50 (2013) 623–637.
- D. Ramadan, M.T.C. McCrudden, A.J. Courtenay, R.F. Donnelly, Enhancement strategies for transdermal drug delivery systems: current trends and applications, *Drug Deliv. Transl. Res.* (2021) 1–34.
- A.D. Permana, F. Nainu, K. Moffatt, E. Larrañeta, R.F. Donnelly, Recent advances in combination of microneedles and nanomedicines for lymphatic targeted drug delivery, *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.* (2021) 1–22.
- A.D. Permana, I.A. Tekko, M.T.C. McCrudden, Q.K. Anjani, D. Ramadan, H. O. McCarthy, R.F. Donnelly, Solid lipid nanoparticle-based dissolving microneedles: a promising intradermal lymph targeting drug delivery system with potential for enhanced treatment of lymphatic filariasis, *J. Control. Release* 316 (2019) 34–52.
- A.D. Permana, A.J. Paredes, F. Volpe-Zanutto, Q.K. Anjani, E. Utomo, R. F. Donnelly, Dissolving microneedle-mediated dermal delivery of itraconazole nanocrystals for improved treatment of cutaneous candidiasis, *Eur. J. Pharm. Biopharm.* 154 (2020) 50–61, <https://doi.org/10.1016/j.ejpb.2020.06.025>.
- A.D. Permana, M.T.C. McCrudden, R.F. Donnelly, Enhanced intradermal delivery of nanosuspensions of antifilarial drugs using dissolving microneedles: a proof of concept study, *Pharmaceutics* 11 (2019) 346.
- M.M. Badran, J. Kuntsche, A. Fahr, Skin penetration enhancement by a microneedle device (Dermaroller®) in vitro: dependency on needle size and applied formulation, *Eur. J. Pharm. Sci.* 36 (2009) 511–523, <https://doi.org/10.1016/j.ejps.2008.12.008>.
- A. Ahad, A.A. Al-Saleh, A.M. Al-Mohizea, F.I. Al-Jenoobi, M. Raish, A.E.B. Yassin, M.A. Alam, Pharmacodynamic study of eprosartan mesylate-loaded transfersomes Carbopol® gel under Dermaroller® on rats with methyl prednisolone acetate-induced hypertension, *Biomed. Pharmacother.* 89 (2017) 177–184, <https://doi.org/10.1016/j.biopha.2017.01.164>.
- K.S. Ahmed, X. Shan, J. Mao, L. Qiu, J. Chen, Derma roller® microneedles-mediated transdermal delivery of doxorubicin and celecoxib co-loaded liposomes for enhancing the anticancer effect, *Mater. Sci. Eng. C* 99 (2019) 1448–1458, <https://doi.org/10.1016/j.msec.2019.02.095>.
- A.D. Permana, E. Utomo, M.R. Pratama, M.N. Amir, Q.K. Anjani, S.A. Mardikasari, S. Sumarheni, A. Himawan, A. Arjuna, U. Usmanengsi, R.F. Donnelly, Bioadhesive-thermosensitive in situ vaginal gel of the gel flake-solid dispersion of itraconazole for enhanced antifungal activity in the treatment of vaginal candidiasis, *ACS Appl. Mater. Interfaces* 13 (2021) 18128–18141, <https://doi.org/10.1021/acsaami.1c03422>.
- J. Chen, R. Zhou, L. Li, B. Li, X. Zhang, J. Su, Mechanical, rheological and release behaviors of a poloxamer 407/poloxamer 188/carbopol 940 thermosensitive composite hydrogel, *Molecules* 18 (2013) 12415–12425, <https://doi.org/10.3390/molecules181012415>.
- C.K. Enggi, H.T. Isa, S. Sulistiawati, K.A.R. Ardika, S. Wijaya, R.M. Asri, S. A. Mardikasari, R.F. Donnelly, A.D. Permana, Development of thermosensitive and mucoadhesive gels of cabotegravir for enhanced permeation and retention profiles in vaginal tissue: a proof of concept study, *Int. J. Pharm.* 609 (2021), 121182, <https://doi.org/10.1016/j.ijpharm.2021.121182>.
- A.D. Permana, R. Nurul, P. Layadi, A. Himawan, N. Juniarti, Q. Kurnia, E. Utomo, S. Aulia, A. Arjuna, R.F. Donnelly, Thermosensitive and mucoadhesive in situ ocular gel for effective local delivery and antifungal activity of itraconazole nanocrystal in the treatment of fungal keratitis, *Int. J. Pharm.* 602 (2021), 120623.
- L. Rahman, R.S. Lembang, S. Lallo, S.R. Handayani, Usmanengsi, A.D. Permana, Bioadhesive dermal patch as promising approach for improved antibacterial activity of bioactive compound of Zingiber cassumunar Roxb in ex vivo *Staphylococcus aureus* skin infection model, *J. Drug Deliv. Sci. Technol.* 63 (2021), 102522, <https://doi.org/10.1016/j.jddst.2021.102522>.
- S. Khan, M.U. Minhas, I.A. Tekko, R.F. Donnelly, R.R.S. Thakur, Evaluation of microneedles-assisted in situ depot forming poloxamer gels for sustained transdermal drug delivery, *Drug Deliv. Transl. Res.* 9 (2019) 764–782, <https://doi.org/10.1007/s13346-019-00617-2>.
- S.S. Deshkar, V.K. Palve, Formulation and development of thermosensitive cyclodextrin-based in situ gel of voriconazole for vaginal delivery, *J. Drug Deliv. Sci. Technol.* 49 (2019) 277–285, <https://doi.org/10.1016/j.jddst.2018.11.023>.
- F. Mo, J. Ma, X. Yang, P. Zhang, Q. Li, J. Zhang, In vitro and in vivo effects of the combination of myricetin and miconazole nitrate incorporated to thermosensitive hydrogels, on *C. albicans* biofilms, *Phytomedicine* 71 (2020), 153223, <https://doi.org/10.1016/j.phymed.2020.153223>.
- K. Al Khateb, E.K. Ozhmukhametova, M.N. Mussin, S.K. Seilkhanov, T. K. Rakhypbekov, W.M. Lau, V.V. Khutoryanskiy, In situ gelling systems based on Pluronic F127/Pluronic F68 formulations for ocular drug delivery, *Int. J. Pharm.* 502 (2016) 70–79, <https://doi.org/10.1016/j.ijpharm.2016.02.027>.
- S. Shaarani, S.S. Hamid, N.H.M. Kaus, The Influence of pluronic F68 and F127 nanocarrier on physicochemical properties, in vitro release, and antiproliferative activity of thymoquinone drug, *Pharmacogn. Res.* 9 (2017) 12–20, <https://doi.org/10.4103/0974-8490.199774>.
- A. Himawan, N.J.N. Djide, S.A. Mardikasari, R.N. Utami, A. Arjuna, R.F. Donnelly, A.D. Permana, A novel in vitro approach to investigate the effect of food intake on release profile of valsartan in solid dispersion-floating gel in-situ delivery system, *Eur. J. Pharm. Sci.* 168 (2022), 106057, <https://doi.org/10.1016/j.ejps.2021.106057>.
- S. Walfish, Analytical methods: a statistical perspective on the ICH Q2A and Q2B guidelines for validation of analytical methods, *BioPharm. Int.* (2007) 1–6.
- A. Ahad, A.A. Al-Saleh, A.M. Al-Mohizea, F.I. Al-Jenoobi, M. Raish, A.E.B. Yassin, M.A. Alam, Pharmacodynamic study of eprosartan mesylate-loaded transfersomes Carbopol® gel under Dermaroller® on rats with methyl prednisolone acetate-induced hypertension, *Biomed. Pharmacother.* 89 (2017) 177–184.
- R.F. Donnelly, T.R.R. Singh, A.Z. Alkilani, M.T.C. McCrudden, S. O'Neill, C. O'Mahony, K. Armstrong, N. McLoone, P. Kole, A.D. Woolfson, Hydrogel-forming microneedle arrays exhibit antimicrobial properties: Potential for enhanced patient safety, *Int. J. Pharm.* 451 (2013) 76–91, <https://doi.org/10.1016/j.ijpharm.2013.04.045>.
- G. Albon, Safety and effectiveness of an automated microneedling device in improving the signs of aging skin, *J. Clin. Aesthet. Dermatol.* 11 (2018) 29–34.